

GENOME-WIDE SELECTION SIGNATURES IN PINZGAU CATTLE

Radovan Kasarda, Nina Moravčíková, Anna Trakovická, Gábor Mészáros, Ondrej Kadlečík

ABSTRACT

The aim of this study was to identify the evidence of recent selection based on estimation of the integrated Haplotype Score (iHS), population differentiation index (F_{ST}) and characterize affected regions near QTL associated with traits under strong selection in Pinzgau cattle. In total 21 Austrian and 19 Slovak purebred bulls genotyped with Illumina bovineHD and bovineSNP50 BeadChip were used to identify genomic regions under selection. Only autosomal loci with call rate higher than 90%, minor allele frequency higher than 0.01 and Hardy-Weinberg equilibrium limit of 0.001 were included in the subsequent analyses of selection sweeps presence. The final dataset was consisted from 30538 SNPs with 81.86 kb average adjacent SNPs spacing. The iHS score were averaged into non-overlapping 500 kb segments across the genome. The F_{ST} values were also plotted against genome position based on sliding windows approach and averaged over 8 consecutive SNPs. Based on integrated Haplotype Score evaluation only 7 regions with iHS score higher than 1.7 was found. The average iHS score observed for each adjacent syntenic regions indicated slight effect of recent selection in analysed group of Pinzgau bulls. The level of genetic differentiation between Austrian and Slovak bulls estimated based on F_{ST} index was low. Only 24% of F_{ST} values calculated for each SNP was greater than 0.01. By using sliding windows approach was found that 5% of analysed windows had higher value than 0.01. Our results indicated use of similar selection scheme in breeding programs of Slovak and Austrian Pinzgau bulls. The evidence for genome-wide association between signatures of selection and regions affecting complex traits such as milk production was insignificant, because the loci in segments identified as affected by selection were very distant from each other. Identification of genomic regions that may be under pressure of selection for phenotypic traits to better understanding of the relationship between genotype and phenotype is one of the challenges for livestock genetics.

Keywords: bovine SNP50 BeadChip; F_{ST} ; iHS score; Pinzgau cattle; selection sweep

INTRODUCTION

Genome-wide screening of single nucleotide polymorphisms (SNPs) can improve the understanding of the connection between genotype and phenotype changes resulting from the formation of modern livestock breeds. The analysis of a large number of SNPs across the genome will reveal aspects of the population genetic structure, including evidence of adaptive selection across the genome (Barendse et al., 2009). Variations identified within the genome of cattle breeds have been primarily caused by human selection during the processes of domestication and subsequent breed formation. Domestication greatly changed the morphological and behavioral characteristics of cattle and with breed formation and selection programmes for improving the production traits allowed the formation of very diverse breeds (De Simoni Gouveia et al., 2014).

The explanation and identification of selection signatures can provide not only basic knowledge about evolutionary changes which shaped the genome but also can be very perspective for identifying domestication-related loci that ultimately may help to further genetically improve of economically important traits (De Simoni Gouveia et al., 2014; Qanbari et al., 2014). Much of the variation across the genetically diverse ancestral population was either lost

due to limited numbers of animals within the areas of domestication or was divided into the subpopulations that were later recognized as distinct breeds. The strong selection to fix favourable mutations underlying domestication and formation of each breed created selective sweeps in which the variation was also lost (Ramey et al., 2013). If the mutation was recent and the selection is strong all alleles under positive selection will increase in frequency by producing selective sweep or selection signature. For neutral mutation, this will take many generations until the mutated allele has reached a high population frequency through drift. Where the loci selection is slight or the mutation is old little evidence of this selection may be left in the genome (Qanbari et al., 2010a; Kemper et al., 2014).

The evaluation of genes underlying phenotypic variation can be prepared based on two approaches: firstly from phenotype to genome that is carried out by linkage disequilibrium based association mapping and may involve positional cloning of QTL or by targeting particular candidate genes identified based on homology to known genes and secondly from genome to phenotype that includes the statistical estimation of genomic data to identify likely targets of past selection. The elimination of standing variation in regions linked to a recently fixed

beneficial mutation is known as a “selective sweep” (Moradi et al., 2012; Qanbari et al., 2010b, Utsunomiya et al., 2013). Several methods were used to assess the evidence of positive selection including the analysis of population differences using F_{ST} , the analysis of ancestral states in connection with extended haplotype homozygosity (EHH) of derived alleles (iHS) and the modeling of distribution of allele frequencies along chromosome under expectation of a selective sweep analyses by composite likelihood ratio (CLR) (Barendse et al., 2009). For detection of recent selection via selective sweeps identification Sabeti et al. (2002) suggested basic concept resulting from the extended haplotype homozygosity statistic. Sabeti et al. (2002) proposed the use of the contrast of EHH statistic of one core haplotype vs. other haplotypes in the same position if the selection carries an allele on a specific haplotype to higher frequency faster than the rate at which it is broken down by recombination when the high frequency will be longer than expected under neutrality (Qanbari et al., 2010b; Simianer et al., 2010). The integrated Haplotype Score as an extension of EHH statistic was proposed by Voight et al. (2006). This method is based on the comparison of EHH between derived alleles and ancestral alleles present in the wild ancestor or outgroups within population (Simianer et al., 2010). The allele frequency differences between breeds or populations in a segment of genome caused by different histories between them can be analysed by F_{ST} . Fixation index F_{ST} can be interpreted also as a measure of gene frequencies dispersion among groups relative to the variation expected in the population from which such groups was derived (Kemper et al., 2014).

The aim of our study was to identify the signatures of strong and recent selection based on estimation of the integrated Haplotype Score and population differentiation index and characterise genomic regions which have been subjected to selective sweeps.

MATERIAL AND METHODOLOGY

In this study three data sets of animals were used to detect signatures of recent selection in Pinzgau cattle. In total 21 Austrian and 19 Slovak purebred bulls registered by their breed association were genotyped using Illumina BovineHD and Illumina BovineSNP50 BeadChip. Dataset of 21 Austrian sires was builded with the aim of having common ancestors or being related to Slovak ones. The detailed description of sample size and data source for each set can be found in Table 1. The 36393 SNPs

common to applied Illumina genotyping arrays were retained in reduced panel of SNPs. Markers assigned to unmaped regions or with unknown chromosomal position according to the latest bovine genome assembly (Btau 4.0) and SNPs positioned to sex chromosomes were removed (634). Quality control of data was carried out according to Purcell et al. (2007). Autosomal loci with call rate <90%, minor allele frequency <0.01 and Hardy-Weinberg equilibrium limit of 0.001 were excluded from subsequent analyses (5221). The evidence of positive selection was evaluated based on two approaches: integrated Haplotype Score (iHS) statistic and Wright’s fixation index (F_{ST}) measure.

The analysis of selection sweeps using iHS statistic is based on haplotype frequencies as specified Voight et al. (2006). The haplotypes were reconstructed for each autosome using default parameters according to Scheet and Stephens (2006). The iHS statistic evaluate the extent of local linkage disequilibrium which is partitioned into haplotypes positioned upon and loci that carry the ancestral versus the derived allele. The iHS score reflects the structure of haplotype and essentially indicates unusually long haplotypes carrying the ancestrall and derived allele (Qanbari et al., 2011). The set of ancestrall alleles resulting from research of Matukumalli et al. (2009) was used in our study. In iHS statistic each loci is treated as core SNP and the test begins with calculation of extended Haplotype Homozygosity for each core SNP. If SNPs are biallelic loci, then each core SNP can be ancestral or derived. This integrated EHH (iHH) (summed over both directions away from the core SNP) is denoted iHH_A or iHH_D , depending on whether it is computed for the ancestral or derived core allele (Qanbari et al., 2011; De Simoni Gouveia et al., 2014). According Voight et al. (2006) the iHS score is described as within population score for the ratio between iHH_A and iHH_D :

$$iHS = \ln \frac{(iHH_A)}{(iHH_D)}$$

The negative iHS values indicate greater homozygosity outlying the ancestral allele and positive values denote greater homozygosity outlying the derived allele. The iHS within analysed population was evaluated using the *rehh* package that is incorporated in R software (Gautier and Vitalis, 2012). Subsequently, the iHS values were averaged in genome-wide non-overlapping 500 kb windows.

Table 1 Description of used sample and genotyping array.

Breed	Country	Sample (n)	SNPs on array	Production type
Pinzgau	Austria	17	700k	
Pinzgau	Austria	4	54001	Dual-purpose
Pinzgau	Slovakia	19	54609	

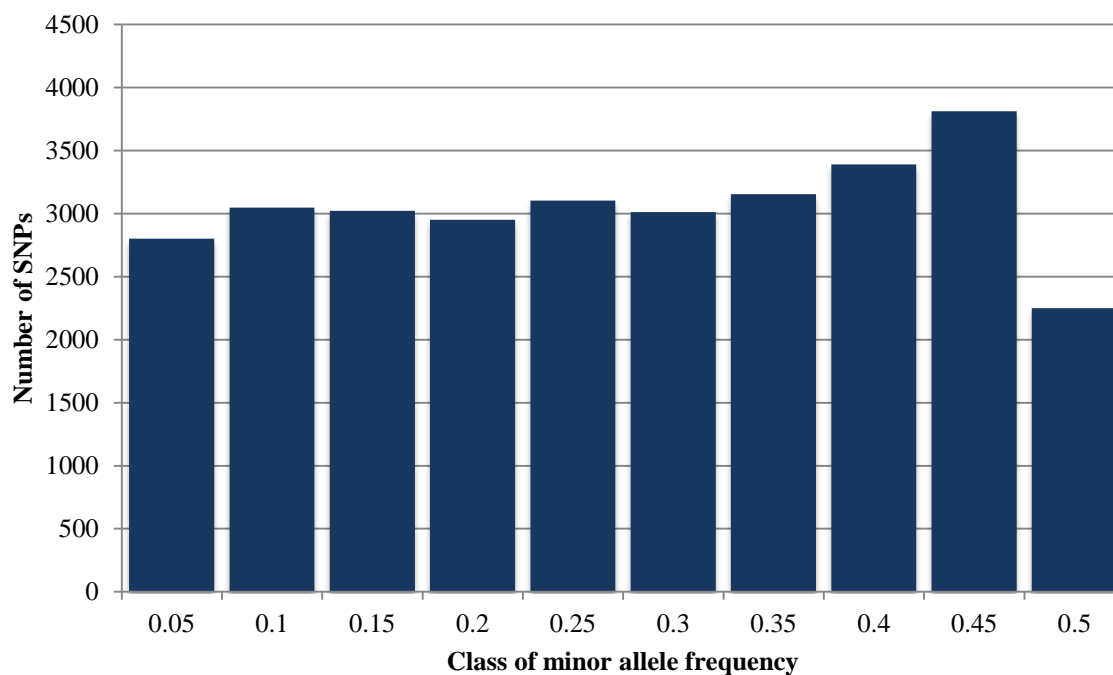


Figure 1 Distribution of MAF across genome.

The genome-wide pattern of selection signatures were estimated also by calculating the basic form of Wright’s F_{ST} fixation index corrected by **Weir and Cockerham (1984)** at each syntenic locus and visualised using SNP & Variation Suite v8.x (**Golden Helix, Inc., Bozeman, MT, www.goldenhelix.com**). The F_{ST} index describing the degree of genetic differentiation between subpopulations can theoretically range from 0 to 1, but it is also possible to assume negative values (**Akey et al., 2002**). Selection signatures can be recognized when adjacent SNPs all show high F_{ST} (**Weir et al., 2005**), due to the hitch-hiking effect (**Maynard-Smith et al., 1974**), implying divergent selection between breeds, or where adjacent SNPs all show low F_{ST} , implying balancing selection between breeds. Smoothing, where a moving average of a certain number of markers is taken, is a method of looking for regions where selection is apparent over multiple markers, rather than one-off high values (**Barendse et al., 2009; Moradi et al., 2012**). The F_{ST} values were also evaluated against genome position based on sliding windows approach and averaged over 8 consecutive SNPs.

RESULTS

The dataset consisting of the total 30538 autosomal SNPs that passed the filtering criteria have been used to identify genomic regions in Pinzgau bulls that may be influenced by recent selection. This subset of loci covered 25084.85 Mbp of the genome with 81.86 kb average adjacent SNPs spacing. The distribution of minor allele frequency (MAF) across the panel of loci was not uniform (Figure 1). The average value of MAFs (0.24 ± 0.14) was comparable with other dual-purpose cattle breeds (**Edel et al., 2011; Su et al., 2012; Mancini et al., 2014**). Across all analysed individuals the average heterozygosity at level 0.34 ± 0.01 was observed.

Two approaches were used for evidence of recent selection. Firstly, the iHS statistic was applied on dataset to detect selection sweeps. The iHS score was calculated for each SNPs and then averaged into non-overlapping 500 kb segments across the genome. The size of sliding windows was chosen based on sufficient number of SNPs for each segment. Genomic regions were considered as recently selected when the iHS score of multiple loci located within 0.5 Mb was greater than 1.7. In total

Table 2 Detected autosome segments identified as regions under selection.

Region	BTA	Position (Mb)	Number of SNPs	iHS value	Closest gene to max
1	1	65.66 – 65.99	3	1.70	HGD
2	5	55.58	1	1.73	LOC101904412
3	7	12.83	1	2.24	DCAF15
4	9	72.11 – 72.31	2	1.70	EYA4
5	15	52.03 – 52.47	7	1.86	NUMA1
6	18	22.26 – 22.38	2	1.73	FTO
7	19	0.47	1	1.90	CA10

Table 3 Summary of autosome regions identified as affected by positive selection.

Region	BTA	Position (Mb)	Number of SNPs	F _{ST} value
1	1	3.40 – 23.73	3	0.23
2	2	1.32 – 13.28	2	0.25
3	4	57.33 – 72.54	5	0.24
4	5	73.45 – 85.22	3	0.22
5	10	12.71 – 26.17	2	0.23
6	28	5.17 – 11.15	3	0.22

4889 windows involving 23677 iHS values were tested. Table 2 showed summaries of the autosomal regions that displaying significant iHS values. Using iHS statistic was found only few SNPs that can be evaluated as loci under selection. Across genome only 7 windows exceeded the iHS value greater than 1.7. Figure 2 displays the genome-wide plot of iHS values against the genomic position. The observed segments were localized close to the different genes. The average value of iHS score was 0.05 and the highest score (2.24) was identified for region on chromosome 7 with only one observed locus. In this genome location was found the presence of gene encoding DDB1 and CUL4 associated factor 15. Most of SNPs that showed significant iHS values were located on chromosome 15 in genomic region ranged from 52.03 to 52.47 Mb. In this bovine autosomal region is located the nuclear mitotic apparatus protein 1 (NUMA1) gene which is gene conserved across different species including human. The NUMA1 gene was tested for evidence of its role in proliferative activity and meiotic cell division (Taimen et al., 2004). The region on chromosome 18 consisting of only 2 loci was near to the FTO (the fat mass and obesity-associated gene) gene which was significantly associated with carcass traits and meat quality in cattle and pigs (Zhang et al., 2011; Dvořáková et al., 2012). However, the distribution of segments with clustered loci was across autosome non-uniform. The values of iHS

score across autosome segments indicated that the analysed regions which can be affected by recent selection showed no major overlap. The slight signals of recent selection can be caused by the smaller sample size or mainly by the fact that the analysed individuals were genetically related.

Secondly, to estimate genome-wide pattern of positive selection within evaluated population of Pinzgau bulls the F_{ST} index for each SNP was calculated. The level of genomic differentiation was evaluated between two groups based on their origin which can lead to increase of allele frequencies in loci that were potentially affected by positive selection. The higher allele frequencies of these loci can be representative to the differentiation in balancing or directional selection, neutrality or other processes that were used in breeding programs of Austrian and Slovak Pinzgau cattle populations.

Theoretically, the F_{ST} values varied from 0 to 1, when both extremes means the total identity (F_{ST}=0) or differentiation (F_{ST}=1) within analysed populations. The selection signatures could be recognized when adjacent loci all show high F_{ST} due to hitch-hiking effect resulting from divergent selection or when adjacent SNPs all show low F_{ST} resulting from the balancing selection between populations (Qanbari et al., 2011). In our study the autosomal regions were recognized as affected by positive selection when the adjacent SNPs showed F_{ST} values

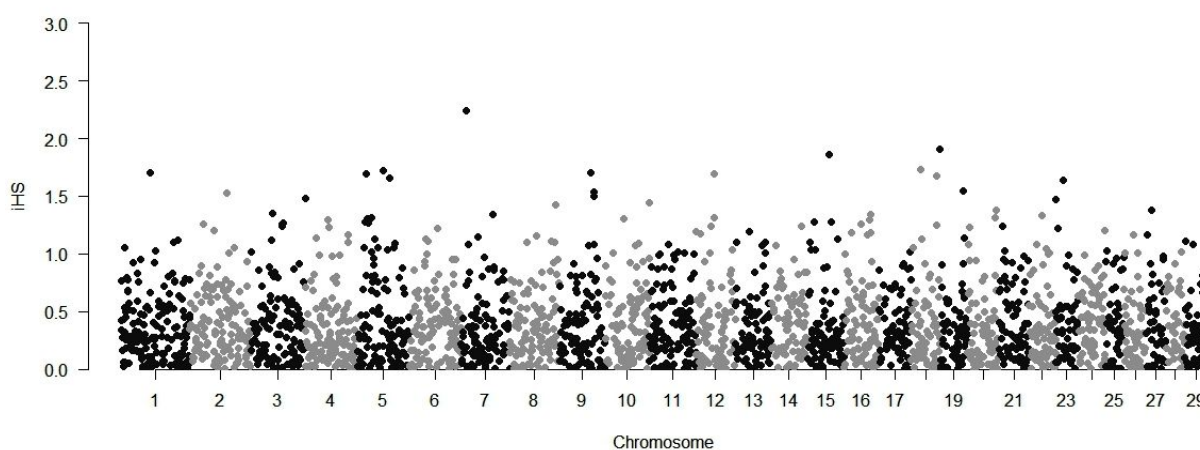


Figure 2 Genome-wide plot of the iHS score averaged for 500 kb.

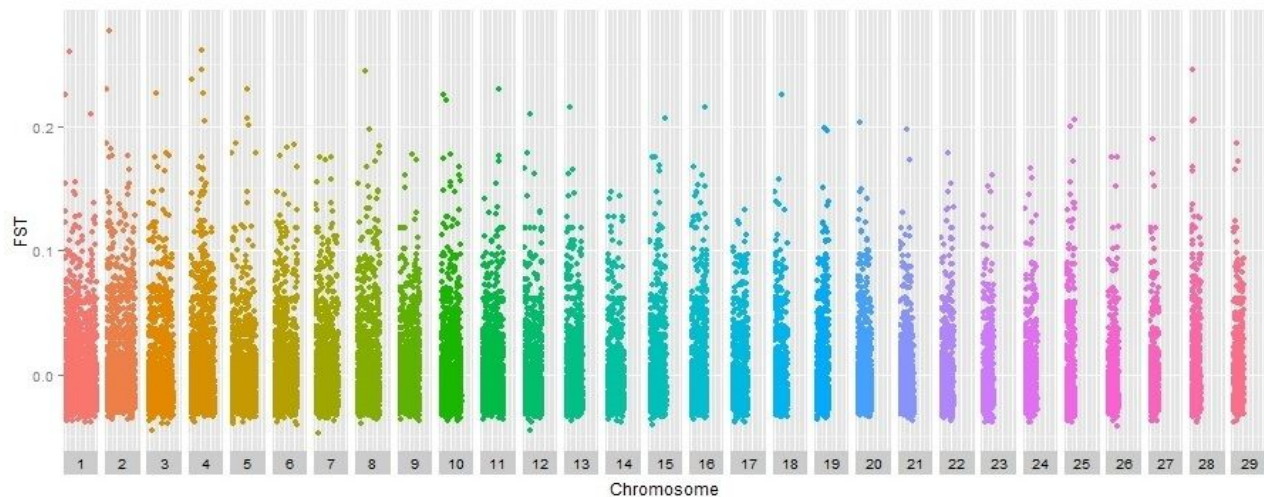


Figure 3 The distribution of F_{ST} values within the autosomes.

higher than 0.20. The observed F_{ST} values ranged from -0.05 to 0.28, with an average values of 0.0005. In total 76.23% of F_{ST} values were lower than 0.01. The highest average F_{ST} was found for BTA 4 (0.004). As indicated in figure 3 only few loci had a tendency to cluster into similar region. The slight signals of positive selection was found in 6 regions localized on chromosomes 1, 2, 4, 5, 10 and 28 (Table 3).

In the next step of F_{ST} estimation the values were averaged over 8-wide SNPs windows within each autosome to determine global pattern of F_{ST} across genome. More than 95% of clusters across all autosomes showed F_{ST} values lower than 0.01. The results indicated unimodal distribution and pretty much uniform scheme of selection in all loci included in analysis. The low level of genetic differentiation between Austrian and Pinzgau bulls detected based on F_{ST} index is also apparent from the similar origin and common ancestors in pedigree data of the analysed Pinzgau bulls.

DISCUSSION

The level of genetic variation among cattle population is a result of both neutral demographic processes, weak but sustained natural selection and strong short-term artificial selection for divergent breeding goals (Qanbari et al., 2011). In our study we used the genome-wide SNP data to detect the evidence of positive selection signals based on the *iHS* and F_{ST} scan in Pinzgau bulls originating from Austria and Slovakia. The results from both analyses indicated low level of genomic differentiation between analysed groups. Based on the *iHS* statistic seven regions that can be potentially affected by recent selection were detected. The total number of SNPs for each region was low. Only one of identified segments was found in genome region associated in previous studies with bovine quantitative trait locus (Table 2). The F_{ST} values averaged in 8-wide windows indicated that the selection programs of Slovak and Austrian bulls are similar and therefore the genomic regions showed only very small differences.

Both of the applied statistics were used successfully in the evaluation of selection signatures in different cattle

populations. Qanbari et al. (2010b) found in population of German Holstein-Friesian cattle segment with an outlier value on chromosome 18 that contains the Sialic acid binding Ig-like lectin 5 gene and the Zink finger protein 577 gene. These genes are considered as candidate for calving ease, longevity and total merit index in Holstein cattle (Cole et al., 2009). Gu et al. (2009) reported the positive selection signatures in the genomic region surrounding muscle related genes. The evidence of strong selection in the region near to the growth hormone gene located on chromosome 20 was found by Flori et al. (2009) in Angus and by Hayes et al. (2009) in Holstein breeds. Simianer et al. (2010) reported the outlier F_{ST} windows for the two regions on chromosome 2 and 5 in the vicinity of ZRANB3, R3HDM1 and WIF1 genes which are known as genes affecting feed efficiency and mammalian mesoderm segregation. In studies of three French dairy cattle breeds published by Van Tassell et al. (2008) and Karim et al. (2011) was found the differentiations in the region located on chromosome 18 which was associated with coat color (MC1R gene) and in the segment on chromosome 14 harbouring the PLAG1 gene that is important for the cattle growth. The platelet-derived growth factor alpha polypeptide (PDGFA) was identified as a potential candidate gene underlying the selective sweep on BTA25 in Simmental cattle and the receptor for this growth factor (PDGFRA) was identified as differentiated among the French dairy breeds (Ramey et al., 2013).

The genome-wide scan for evidence of selection signatures in livestock is one of the many approaches that are available for estimation of genomic diversity due to development of high throughput SNP genotyping arrays. The large observed datasets with high SNPs density provide a much better insight into the biological processes underlying natural and artificial selection of animals. The results of these studies can provide the valuable data for increase of animal selection strategies efficiency and also may help to understanding of biological limits and signals resulting from the high selection pressure of the achievement of breeding goals.

CONCLUSION

The genome-wide scan based on estimation of positive selection signatures using iHS and F_{ST} statistics led to the detection of few regions that were affected by recent selection in Pinzgau cattle. Our results indicated use of similar selection programs in Slovak and Austrian Pinzgau populations. The conditions that would result in a clear evidence of selection signatures were rare. The response to the selection resulted from the small allele frequency changes in many loci that were polymorphic before start of selection in population. The results show low level of genetic differentiation or high genetic relatedness between analysed Austrian and Slovak Pinzgau bulls what is due to the fact that bulls from both populations had common ancestors. Pinzgau cattle are recognized as producer of food resources of specific quality due to its mountaineous origin. Observed results confirmed previously stated assumption of common genetic pool of Slovak and Austrian populations and indicated importance of both populations in preservation and utilization of Pinzgau cattle.

REFERENCES

- Akey, J. M., Zhang, G., Zhang, K., Jin, L., Shriver, M. D. 2002. Interrogating a high-density SNP map for signatures of natural selection. *Genome Research*, vol. 12, p. 1805-1814. <http://dx.doi.org/10.1101/gr.631202>
- Barendse, W., Harrison, B., Bunch, R., Thomas, M., Turner, L. 2009. Genome wide signatures of positive selection: The comparison of independent samples and the identification of regions associated to traits. *BMC Genomics*, vol. 10, p. 178. <http://dx.doi.org/10.1186/1471-2164-10-178>
- Cole, J. B., VanRaden, P. M., O'Connell, J. R., Van Tassell, C. P., Sonstegard, T. S., Schnabel, R. D., Taylor, J. F., Wiggans, G. R. 2009. Distribution and location of genetic effects for dairy traits. *Journal of Dairy Science*, vol. 92, p. 2931-2946. <http://dx.doi.org/10.3168/jds.2008-1762>
- de Simoni Gouveia, J. J., da Silva, M. V., Paiva, S. R., de Oliveira, S. M. 2014. Identification of selection signatures in livestock species. *Genetics and Molecular Biology*, vol. 37, p. 330-342. <http://dx.doi.org/10.1590/s1415-47572014000300004>
- Dvořáková, V., Bartenschlager, H., Stratil, A., Horák, P., Stupka, R., Cítek, J., Sprysl, M., Hrdlicová, A., Geldermann, H. 2012. Association between polymorphism in FTO gene and growth and carcass traits in pig crosses. *Genetic Selection Evolution*, vol. 17, p. 13. <http://dx.doi.org/10.1186/1297-9686-44-13>
- Edel, C., Schwarzenbacher, H., Hamann, H., Neuner, S., Emmerling, R., Götz, K. U. 2011. The German-Austrian Genomic Evaluation System for Fleckvieh (Simmental) Cattle. [online] 2015-03-24. [cit. 2015-03-24] Available at: <https://journal.interbull.org/index.php/ib/article/viewFile/1209/1319>.
- Flori, L., Fritz, S., Jaffrézic, F., Boussaha, M., Gut, I., Heath, S., Foulley, J. L., Gautier, M. 2009. The genome response to artificial selection: a case study in dairy cattle. *PLoS One*, vol. 4, p. e6595. <http://dx.doi.org/10.1371/journal.pone.0006595>
- Gautier, M., Vitalis, R. 2012. rehh: an R package to detect footprints of selection in genome-wide SNP data from haplotype structure. *Bioinformatics*, vol. 28, p. 1176-1177. <http://dx.doi.org/10.1093/bioinformatics/bts115>
- Gu, J., Orr, N., Park, S. D., Katz, L. M., Sulimova, G., MacHugh, D. E., Hill, E. W. 2009. A Genome Scan for Positive Selection in Thoroughbred Horses. *PLoS One*, vol. 4, p. e5767. <http://dx.doi.org/10.1371/journal.pone.0005767>
- Hayes, B. J., Chamberlain, A. J., Maceachern, S., Savin, K., McPartlan, H., MacLeod, I., Sethuraman, L., Goddard, M. E. 2009. A genome map of divergent artificial selection between *Bos taurus* dairy cattle and *Bos taurus* beef cattle. *Animal Genetics*, vol. 40, vol. 176-184. [PMid:19067671](https://pubmed.ncbi.nlm.nih.gov/19067671/)
- Karim, L., Takeda, H., Lin, L., Druet, T., Arias, J. A. C., Baurain, D., Cambisano, N., Davis, S. R., Farnir, F., Grisart, B., Harris, B. L., Keehan, M. D., Littlejohn, M. D., Spelman, R. J., Georges, M., Coppieters, W. 2011. Variants modulating the expression of a chromosome domain encompassing PLAG1 influence bovine stature. *Nature Genetics*, vol. 43, p. 405-413. [PMid:21516082](https://pubmed.ncbi.nlm.nih.gov/21516082/)
- Kemper, K. E., Saxton, S. J., Bolormaa, S., Hayes, B. J., Goddard, M. E. 2014. Selection for complex traits leaves little or no classic signatures of selection. *BMC Genomics*, vol. 326, p. 246. <http://dx.doi.org/10.1186/1471-2164-15-246>
- Mancini, G., Gargani, M., Chillemi, G., Nicolazzi, E. L., Marsan, P. A., Valentini, A., Pariset, L. 2014. Signatures of selection in five Italian cattle breeds detected by a 54K SNP panel. *Molecular Biology Reports*, vol. 41, p. 957-965. <http://dx.doi.org/10.1007/s11033-013-2940-5>
- Matukumalli, L. K., Lawley, C. T., Schnabel, R. D., Taylor, J. F., Allan, M. F., Heaton, M. P., O'Connell, J., Moore, S. S., Smith, T. P., Sonstegard, T. S., Van Tassell, C. P. 2009. Development and Characterization of a High Density SNP Genotyping Assay for Cattle. *PLoS One*, vol. 4, p. e5350. <http://dx.doi.org/10.1371/journal.pone.0005350>
- Maynard-Smith, J., Haigh, J. 1974. The hitch-hiking effect of a favourable gene. *Genetical Research*, vol. 23, p. 23-35. <http://dx.doi.org/10.1017/s0016672300014634>
- Moradi, M. H., Nejati-Javaremi, A., Moradi-Shahrbabak, M., Dods, K. G., McEwan, J. C. 2012. Genomic scan of selective sweeps in thin and fat tail sheep breeds for identifying of candidate regions associated with fat deposition. *BMC Genetics*, vol. 13, p. 10. <http://dx.doi.org/10.1186/1471-2156-13-10>
- Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M. A. R., Bender, D., Maller, J., Sklar, P., de Bakker, P. I. W., Daly, M. J., Sham, P. C. 2007. PLINK: a toolset for whole-genome association and population-based linkage analysis. *American Journal of Human Genetics*, vol. 81, p. 559-575. <http://dx.doi.org/10.3410/f.1162373.622875>
- Qanbari, S., Gianola, D., Hayes, B., Schenkel, F., Miller, S., Moore, S., Thaller, G., Simianer, H. 2011. Application of site and haplotype-frequency based approaches for detecting selection signatures in cattle. *BMC Genomics*, vol. 12, p. 318. <http://dx.doi.org/10.1186/1471-2164-12-318>
- Qanbari, S., Pausch, H., Jansen, S., Somel, M., Strom, T. M., Fries, R., Nielsen, R., Simianer, H. 2014. Classic Selective Sweeps Revealed by Massive Sequencing in Cattle. *PLoS Genetics*, vol. 10, p. e1004148. <http://dx.doi.org/10.1371/journal.pgen.1004148>
- Qanbari, S., Pimentel, E. C. G., Tetens, J., Thaller, G., Lichtner, P., Sharifi, A.R., Simianer, H. 2010a. A Genome-Wide Scan for Signatures of Recent Selection in Holstein Cattle. *Animal Genetics*, vol. 41, p. 377-389. [PMid:20096028](https://pubmed.ncbi.nlm.nih.gov/20096028/)
- Qanbari, S., Pimentel, E. C. G., Tetens, J., Thaller, G., Lichtner, P., Sharifi, A. R., Simianer, H. 2010b. The pattern

of linkage disequilibrium in German Holstein cattle. *Animal Genetics*, vol. 41, p. 346-356. [PMid:20055813](#)

Ramey, H. R., Decker, J. E., McKay, S. D., Rolf, M. M., Schnabel, R. D., Taylor, J. F. 2013. Detection of selective sweeps in cattle using genome-wide SNP data. *BMC Genomics*, vol. 14, p. 382. <http://dx.doi.org/10.1186/1471-2164-14-382>

Sabeti, P. C., Reich, D. E., Higgins, J. M., Levine, H. Z., Richter, D. J., Schaffner, S. F., Gabriel, S. B., Platko, J. V., Patterson, N. J., McDonald, G. J., Ackerman, H. C., Campbell, S. J., Altshuler, D., Cooper, R., Kwiatkowski, D., Ward, R., Lander, E. S. 2002. Detecting recent positive selection in the human genome from haplotype structure. *Nature*, vol. 419, p. 832-837. [PMid:12397357](#)

Scheet, P., Stephens, M. 2006. A fast and flexible statistical model for large-scale population genotype data: Applications to inferring missing genotypes and haplotypic phase. *American Journal of Human Genetics*, vol. 78, p. 629-644. <http://dx.doi.org/10.1086/502802>

Simianer, H., Qanbari, S., Gianola, D. 2010. Detection of selection signatures within and between cattle populations. [online] 2015-03-24. [cit. 2015-03-24] Available at: <http://www.kongressband.de/wcgalp2010/assets/pdf/0618.pdf>

SNP & Variation Suite (Version 8.x) [Software]. Bozeman, MT: Golden Helix, Inc. [cit. 2015-03-24] Available at <http://www.goldenhelix.com>

Su, G., Brøndum, R. F., Ma, P., Guldbrandsen, B., Aamand, G. P., Lund, M. S. 2012. Comparison of genomic predictions using medium density (~54,000) and high density (~777,000) single nucleotide polymorphism markers panel in Nordic Holstein and Red Dairy Cattle populations. *Journal of Dairy Science*, vol. 95, p. 4657-4665. <http://dx.doi.org/10.3168/jds.2012-5379>

Taimen, P., Berghäll, H., Vainionpää, R., Kallajoki, M. 2004. NuMA and nuclear lamins are cleaved during viral infection--inhibition of caspase activity prevents cleavage and rescues HeLa cells from measles virus-induced but not from rhinovirus 1B-induced cell death. *Virology*, vol. 320, p. 85-98. <http://dx.doi.org/10.1016/j.virol.2003.11.026>

Utsunomiya, Y. T., Pérez O'Brien, A. M., Sonstegard, T. S., Van Tassell, C. P., do Carmo, A. S., Mészáros, G., Sölkner, J., Garcia, J. F. 2013. Detecting loci under recent positive selection in dairy and beef cattle by combining different genome-wide scan methods. *PLoS One*, vol. 8, p. e64280. <http://dx.doi.org/10.1371/journal.pone.0064280>

Van Tassell, C. P., Smith, T. P., Matukumalli, L. K., Taylor, J. F., Schnabel, R. D., Lawley, C. T., Haudenschild, C. D., Moore, S. S., Warren, W. C., Sonstegard, T. S. 2008. SNP discovery and allele frequency estimation by deep sequencing of reduced representation libraries. *Nature Methods*, vol. 5, p. 247-252. [PMid:18297082](#)

Voight, B. F., Kudravalli, S., Wen, X., Pritchard, J. K. 2006. A map of recent positive selection in the human genome. *PLoS Biology*, vol. 4, p. e72. [PMid:16494531](#)

Weir, B. S., Cardon, L. R., Anderson, A. D., Nielsen, D. M., Hill, W. G. 2005. Measures of human population structure show heterogeneity among genomic regions. *Genome Research*, vol. 15, p. 1468-1476. <http://dx.doi.org/10.1101/gr.4398405>

Weir, B. S., Cockerham, C. C. 1984. Estimating F-Statistics for the analysis of population structure. *Evolution*, vol. 38, p. 1358-1370. <http://dx.doi.org/10.2307/2408641>

Zhang, B., Zhang, Y., Zhang, L., Wang, J., Li, Z., Chen, H. 2011. Allelic polymorphism detected in the bovine FTO gene. *Molecular Biotechnology*, vol. 49, p. 257-262. <http://dx.doi.org/10.1007/s12033-011-9400-z>

Acknowledgment:

This project was co-financed by the Slovak Research and Development Agency under the contracts no. APVV-0636-11 and APVV-14-0054. Insemas a.s. and SBS a.s. are acknowledged for material support of this research.

Contact address:

Radovan Kasarda, Slovak University of Agriculture in Nitra, Faculty of Agriobiology and Food Resources, Department of Animal Genetics and Breeding Biology, Tr. A. Hlinku 2, 949 76 Nitra, Slovakia, E-mail: RadovanKasarda@uniag.sk.

Nina Moravčíková, Slovak University of Agriculture in Nitra, Faculty of Agriobiology and Food Resources, Department of Animal Genetics and Breeding Biology, Tr. A. Hlinku 2, 949 76 Nitra, Slovakia, E-mail: nina.moravcikova@uniag.sk.

Anna Trakovická, Slovak University of Agriculture in Nitra, Faculty of Agriobiology and Food Resources, Department of Animal Genetics and Breeding Biology, Tr. A. Hlinku 2, 949 76 Nitra, Slovakia, E-mail: anna.trakovicka@uniag.sk.

Gábor Mészáros, University of Natural Resources and Life Sciences, Division of Livestock Sciences, Gregor-Mendel-Straße 33, 1180 Wien Austria, E-mail: gabor.meszaros@boku.ac.at.

Ondrej Kadlecík, Slovak University of Agriculture in Nitra, Faculty of Agriobiology and Food Resources, Department of Animal Genetics and Breeding Biology, Tr. A. Hlinku 2, 949 76 Nitra, Slovakia, E-mail: ondrej.kadlecik@uniag.sk.